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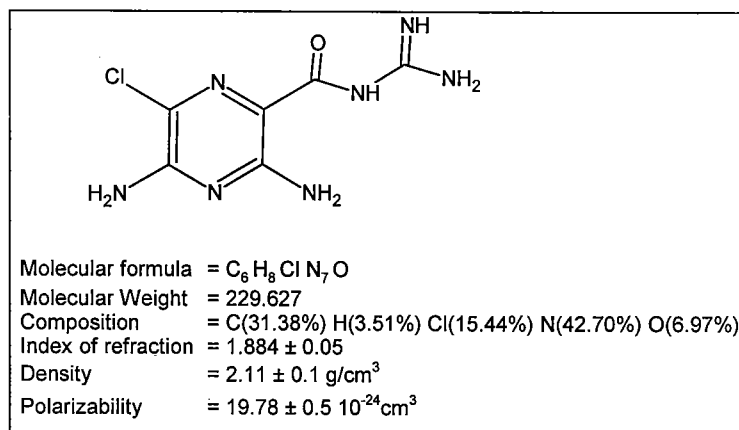
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## Amiloride – Detection and Excretion Study under Conditions of Steroid Screening Procedure

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### INTRODUCTION

Amiloride -3,5 Diamino- N-(aminoiminomethyl) -6-chloropyrazine-carboxamide (Fig.1) is one of the potassium saving diuretics, which are clinically used in cases of potassium deficiency [1]. It is included in the list of banned substances of IOC [2].



**Fig.1.** Structure formula and some theoretical constants of Amiloride.

One of the reasons for more rarely use of Amiloride as diuretic in sport is its effect of a low urinary flow [3]. The problems of detection of Amiloride by the diuretics screening procedure are known. The analytical difficulties result from its structure (very polar molecule containing guanidine and amino groups) which causes a limited solubility in usual organic solvents and problems with derivatization [4-7]. Therefore the aim of our work was to study the behaviour of Amiloride by methods of some screening procedures and to collect data of its excretion.

### EXPERIMENTAL

*1. Excretion study.* An excretion study after an oral application of a therapeutical dose of Amiloride was performed. One volunteer ingested a single dose of 4mg of pure Amiloride. The urine samples were collected up to 70 hours after administration.

## 2. Sample preparation for excretion study.

	To 2ml urine add ISTD Methyltestosterone 40µl (25ppm) apply on XAD -2 column, wash with 2 ml of water elute with 3 ml of methanol, evaporated to dryness
ADD	
	1ml phosphate buffer 0.2M to pH 7.0 25µl β-glucuronidase from E.coli
HYDROLYSIS	
	1 hour at 55°C
ADD	
	100 mg K <sub>2</sub> CO <sub>3</sub> to adjust pH 9.6 5 ml of diethyl ether
EXTRACTION	
	shake for 20 min., centrifuge 5 min. evaporate to dryness
DERIVATIZATION	
	100 µl MSTFA/NH <sub>4</sub> J/ethanethiol 1000:2:6 (v:w:v), 20 min. at 80°C, inject into GC/MS – 1 µl

## 3. GC/MS parameters

- 3) GC/MS:HP 5890/5970, carrier gas He - 1.8ml/min. flow, split ratio 1:10, Column Chrompack CP-Sil 5 CB, 0.25 mm i.D., 0.25 µm film thickness, Temperature program: 150°C/ 1 min./ 13°C/213°C; 2°C/220 °C; 20°C/300°C, Carrier: - 1 ml/min He; Split ratio 1:10; Transfer line – 280 °C; injector – 280 °C. SIM mode for enol-MSTFA derivatives (dwell time: 47 ms) – 293,300,302,308, 310,316, 318, 331, 333, 365, 380, 388,390,403,405,413,415,428,430,476, 478.

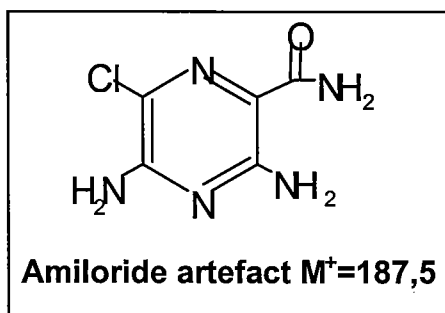
## 4. Extraction study.

- a) Liquid/Liquid extraction - Experiments were performed by 3ml spiked urine with 1 µg/ml Amiloride. Adjust the pH of urine to 9.6 with K<sub>2</sub>CO<sub>3</sub>/NaHCO<sub>3</sub> (2:1), add 5 ml of corresponding solvent and shake for 20 min. Centrifuge, freeze and transfer organic layer to a fresh tube. Dry the sample under a stream of nitrogen in water bath at 40°C. For HPLC analysis samples were dissolved with 200 µl of the mobile phase. The following organic solvents were tested: diethyl ether(DE), ethylacetate (EA) and tert-butylmethyl ether (t-BME).

- b) Solid phase extraction (SPE) on XAD-2 - Spiked urine with Amiloride (1 µg/ml) was applied on a XAD-2 column. The preconditioning of the column and the elution of the sample were carried out as described in the excretion study - sample preparation. The methanol was removed under a stream of nitrogen and the residue was reconstituted in 200 µl of the mobile phase for HPLC.
- c) HPLC conditions – Waters liquid chromatograph, column “NovaPack C18” 150x 39 mm, 4 µm, mobile phase phosphate buffer (pH=2.2), methanol and acetonitrile (85:10.5:4.5) (v/v/v), flow 0.7 ml/min, λ=280 nm.

## RESULTS AND DISCUSSION

The GC property of Amiloride is very poor. One product with a lower molecular mass than that of the underivatized Amiloride was detected by GC/MS analysis. The mass spectrum of this product is shown in Fig.2 a). The retention time (2.26 min), intensive ions at m/e 187,189 and other fragments ions in the MS spectrum support the suggestion that this is an Amiloride artefact with following proposed structure:



Amiloride's methylation under standard conditions for diuretics, in the presence of acetone and  $K_2CO_3$  do not create an appropriate medium for proceeding of this reaction. After derivatization for 5h at 60°C the GC/MS analysis gave rise to signals of the following compounds: Amiloride artefact, monomethylated, dimethylated and trimethylated Amiloride, hexamethylated and heptamethylated Amiloride. The experiment for extractive methylation of Amiloride was not successful. The other attempts for derivatization of Amiloride with MSTFA/MBTFA; MTBSTFA,  $CS_2$  also failed. The spectra of the underivatized Amiloride artefact, the bis- and tetrakis-TMS-derivatives of Amiloride achieved by TMIS, shown in fig. 2.

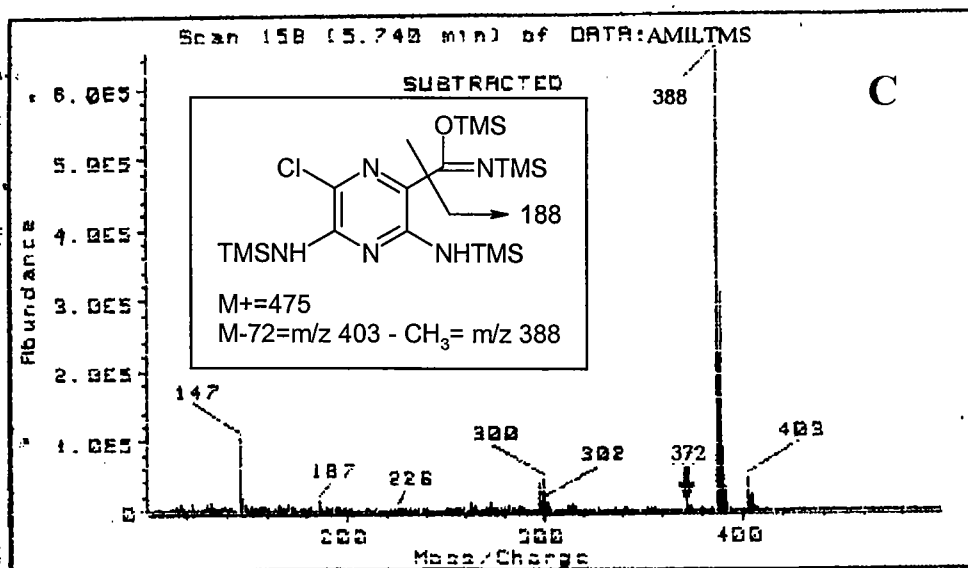
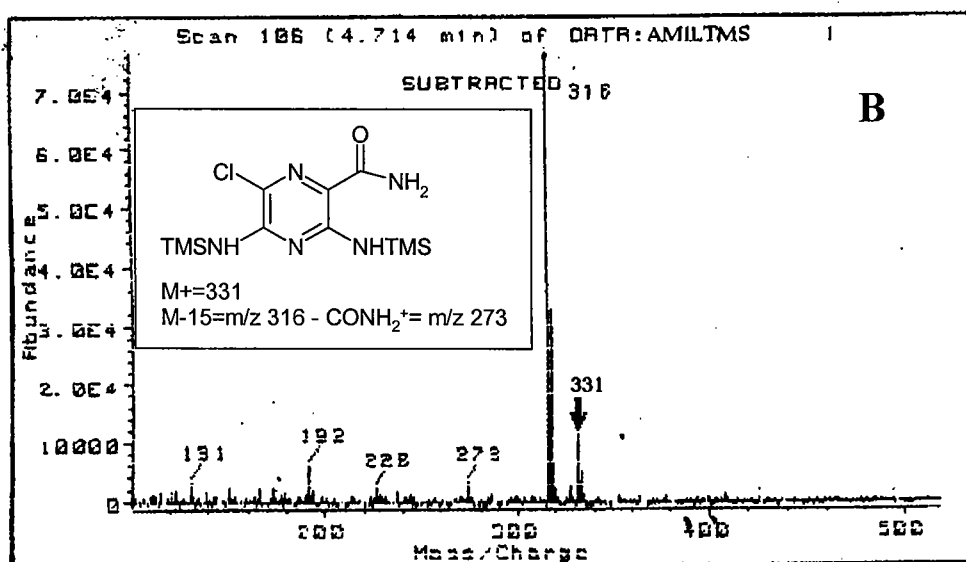
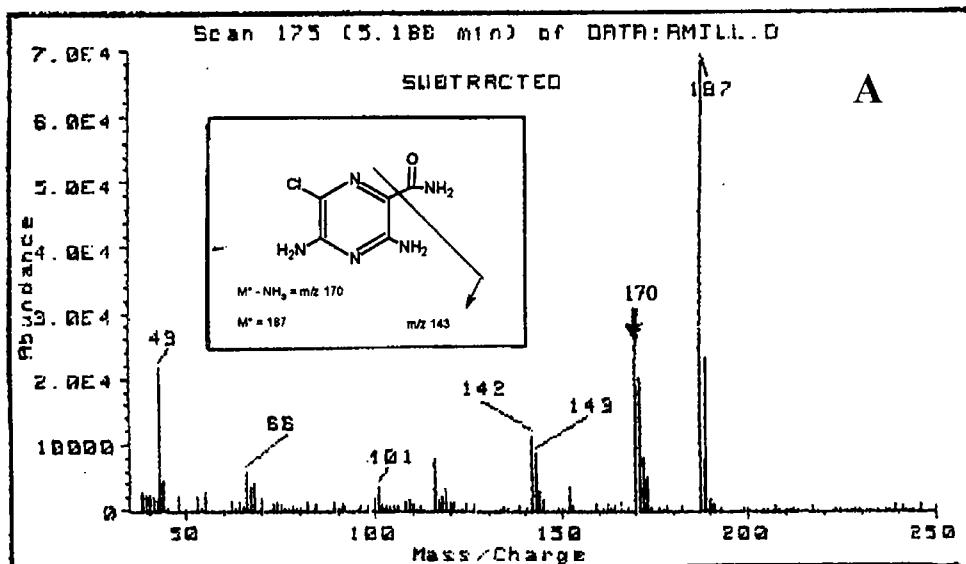


Fig.2. Mass spectra of Amiloride: A) underivatized; B) 2 TMS Artefact; C) 4-TMS Artefact.

2. *Extraction of Amiloride.*- Two chromatographic methods HPLC and GC/MS are used for comparison and determination of the extraction recovery of Amiloride. HPLC analysis of Amiloride was performed without derivatization, and unchanged substance was detected. Enol-TMS derivatization was used in combination with the GC/MS analysis and Amiloride artefact derivatives were detected. In spite of these differences the obtained results are comparable and tendencies are similar (Fig.3). The higher percent of extraction (23%) was obtained when ethylacetate used as extragent for L/L extraction. These results are comparable to those published in literature [8-9]. After applying SPE on XAD-2 column extraction recovery yielded to 10% (by GC/MS) and 12% (by HPLC).

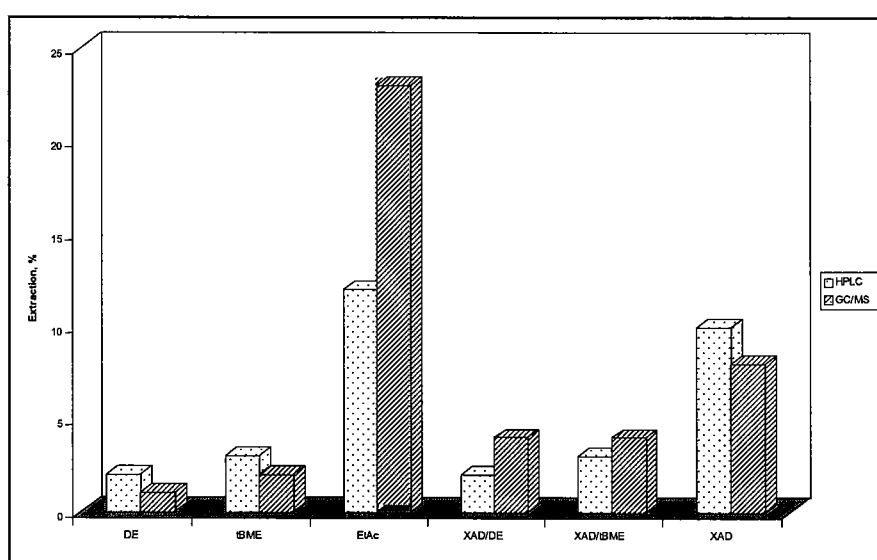


Fig.3. Percents of extraction from spiked urine with 1 µg/ml Amiloride.

3. *Excretion study* – The drugs containing Amiloride are usually combined with other diuretics as Triamterene or Hydrochlorothiazide [1]. This excretion study was performed after oral administration of a single dose (4 mg) of pure substance. Two excretion maxima were obtained – at 3 and 9 hours after administration (Fig.4). In urine samples up to 70 hours after application the Amiloride artefact was detected. Table 1 lists the values of urine parameters and urinary flow up to the first 24 hours after application.

Table 1

Hours after administration	Density, g/cm <sup>3</sup>	pH	Urinary flow, ml
0	1.014	6.8	70
1	1.008	6.7	310
2	1.008	8.2	140

Hours after administration	Density, g/cm <sup>3</sup>	pH	Urinary flow, ml
3	1.018	9.6	100
6	1.019	7.3	100
9	1.024	7.3	80
20	1.018	6.0	300
24	1.018	6.2	150

The limit of detection of the Amiloride artefact by the screening procedure of steroids is 8 ng/ml. The obtained results show the possibility to include Amiloride in the routine screening procedure of anabolic steroids.

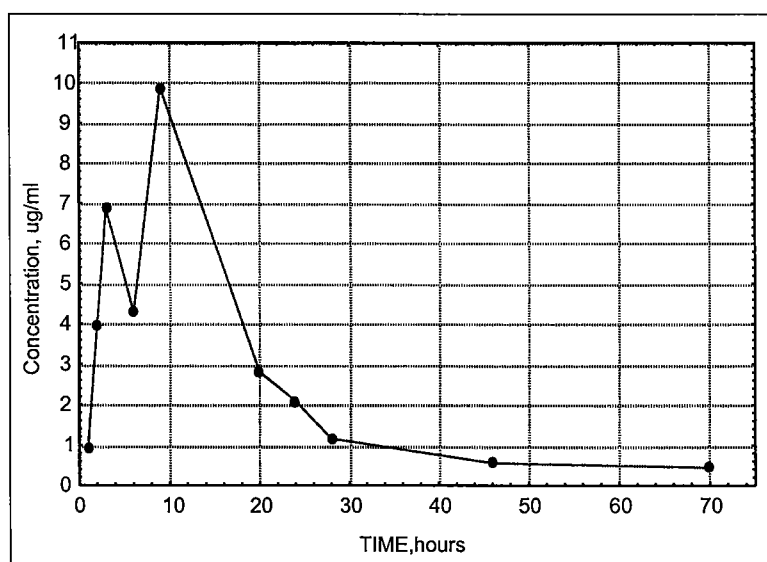


Fig.4. Excretion after oral administration of 4 mg Amiloride.

#### REFERENCES

1. A.Wade (editor), *The Extra Pharmacopoeia*, 29<sup>th</sup> ed., Pharmaceutical Perss, London, 1989,p.543
2. International Olympic Committee, *Prohibited classes of substances and prohibited methods, in Olympic Movement - Anti doping Code and Explanatory Document*, IOC, Lausanne, Switzerland, 2000
3. W.Schanzer, *Official proceedings of International Athletic Foundation World Symposium on doping in sport*, 10-12 May 1987,Fidal, Centro Studi&ricerche,1988
4. R.Ventura, D.Fraise, M. Becchi, O.Paisse, J.Segura, *Journal of chromatography*, (1991), v.562, p.723
5. M.Donike, presented at 9<sup>th</sup> *Workshop on Dope Analysis*, Cologne 1991,17-22 March
6. D.Carreras, C.Imaz, R.Navajas, M.A.Garsia, C.Rodriguez, A.F Rodriguez, R.Cortes, *Journal of Chromatography A*, (1994) v.683, p.195
7. V. Raverdino, *Journal of Chromatography* (1991) v.554, p.125
8. S.F.Cooper, R. Masse,R. Dugal, *Journal of Chromatography*, (1989) v.489, p.65
9. P. Campins-Falco, R. Herraez-Herhahdez, A.Sevilanno-Gabeza, *Journal of Liquid Chromatography*, (1991), v.19, p.3575